

## Pterocidin, a Cytotoxic Compound from the Endophytic *Streptomyces hygroscopicus*

Yasuhiro Igarashi, Shin-suke Miura, Tsuyoshi Fujita, Tamotsu Furumai

Received: September 13, 2005 / Accepted: February 20, 2006

© Japan Antibiotics Research Association

**Abstract** A new cytotoxic compound, pterocidin, was isolated from the endophytic *Streptomyces hygroscopicus* TP-A0451, and the structure was determined on the basis of spectroscopic data. Pterocidin showed cytotoxicity against some human cancer cell lines with IC<sub>50</sub> values of 2.9–7.1 μM.

**Keywords** pterocidin, *Streptomyces*, cytotoxic, δ-lactone

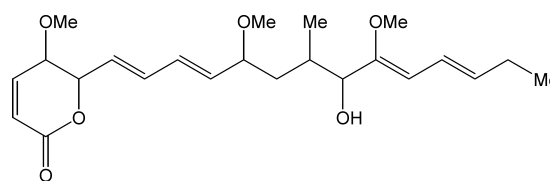
Actinomycetes associated with plants are recognized as an emerging source of novel natural products [1–3]. We have previously reported the isolation of pteridic acids A and B with plant growth promoting activity from the endophytic *Streptomyces hygroscopicus* TP-A0451 [1]. This strain has been so far identified to produce structurally diverse secondary metabolites; an antifungal prenylated indole [2], a sulfonated linear polyene antibiotic with antifungal activity [3], galbonolides A and B [4, 5], elaiophylin [6] and its derivatives, and herbimycins [7] and their hydroquinone congeners. Our continuous search for bioactive compounds from the strain TP-A0451 led to the isolation of pterocidin (**1**), a new cytotoxic compound. We herein describe the isolation and structure elucidation of **1**.

Ten liters of the culture broth of *S. hygroscopicus* TP-A0451 were extracted with 1-butanol, and the extract was fractionated on a silica gel column and further subjected to

C-18 column chromatography to yield pterocidin (**1**, 11 mg) as pale yellow oil.

The molecular formula of **1** was established to be C<sub>23</sub>H<sub>34</sub>O<sub>6</sub> based on <sup>13</sup>C NMR and HRFABMS [*m/z* 429.2260, (M+Na)<sup>+</sup>, Δ +0.7 mmu]. IR absorption of **1** indicated the presence of hydroxyl (3450 cm<sup>-1</sup>) and α,β-unsaturated lactone carbonyl (1720 cm<sup>-1</sup>) groups. The UV absorption [λ<sub>max</sub> (MeOH) 229 (ε 22,900) nm] of **1** was indicative of an α,β-unsaturated carbonyl chromophore. The <sup>1</sup>H and <sup>13</sup>C NMR, DEPT, and HMQC spectra of **1** in CDCl<sub>3</sub> (Table 1) revealed the presence of signals due to one carbonyl carbon, one oxygenated sp<sup>2</sup> quaternary carbon, nine sp<sup>2</sup> methines, four sp<sup>3</sup> methines adjacent to oxygen atoms, one sp<sup>3</sup> methine, two sp<sup>3</sup> methylenes, three methoxy groups, and two methyls. Since six out of 7 unsaturations were accounted for, **1** was inferred to possess one ring.

DQF-COSY spectrum revealed two <sup>1</sup>H-<sup>1</sup>H connectivities from H-2 to H-13 branching H-22 at C-12 and from H-15 to H-19. The HMBC correlations for H-15/C-13 and H-15/C-14 suggested the connectivity between C-13 and C-



**Fig. 1** Structure of pterocidin (**1**).

**Y. Igarashi** (Corresponding author), **S. Miura**, **T. Furumai**: Biotechnology Research Center, Toyama Prefectural University, 5180 Kurokawa, Kosugi, Toyama 939-0398, Japan, E-mail: yas@pu-toyama.ac.jp

**T. Fujita**: Suntory Institute for Bioorganic Research, 1-1-1 Wakayamadai, Shimamoto, Mishima, Osaka 618-8503, Japan

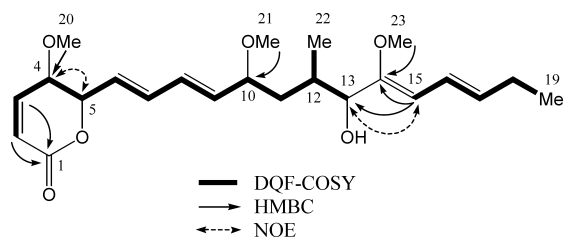
**Table 1** Physico-chemical properties of pterocidin

Appearance	Pale yellow oil
$[\alpha]_D^{20}$	$-27.7^\circ$ ( $c$ 0.46, $\text{CHCl}_3$ )
HRFAB-MS	
Found	429.2260 ( $\text{M}+\text{Na}$ ) <sup>+</sup>
Calcd.	429.2253 (for $\text{C}_{23}\text{H}_{34}\text{O}_6\text{Na}$ )
Molecular formula	$\text{C}_{23}\text{H}_{34}\text{O}_6$
UV (MeOH) $\lambda_{\text{max}}$ nm ( $\epsilon$ )	229 (22,900), 275 (2,500)
IR (neat) $\nu_{\text{max}}$ $\text{cm}^{-1}$	3450, 1720

**Table 2** NMR assignment for pterocidin ( $\text{CDCl}_3$ )

Position	$\delta^{13}\text{C}$	$\delta^1\text{H}$ (mult., $J$ in Hz)
1	162.6	
2	123.3	6.16 (dd, 10.0, 0.7)
3	143.1	6.94 (dd, 10.0, 4.4)
4	71.2	4.02 (ddd, 4.4, 4.2, 0.7)
5	79.8	4.98 (ddd, 6.8, 4.2, 1.0)
6	125.8	5.89 (dd, 15.4, 6.8)
7	134.1	6.44 (ddd, 15.4, 10.5, 1.0)
8	131.4	6.24 (dd, 15.4, 10.5)
9	135.9	5.61 (dd, 15.4, 7.8)
10	79.7	3.73 (m)
11	39.6	1.60 (m), 1.66 (m)
12	33.1	1.96 (m)
13	74.5	4.06 (br d, 3.4)
14	155.3	
15	111.8	5.58 (d, 11.0)
16	122.4	6.34 (ddt, 15.4, 11.0, 1.5)
17	134.8	5.67 (dt, 15.4, 6.6)
18	26.0	2.10 (m)
19	13.7	1.02 (t, 6.8)
20	57.2	3.43 (s)
21	56.3	3.27 (s)
22	13.6	0.89 (d, 6.8)
23	60.0	3.70 (s)

14. The NOE between H-13 and H-15 suggested the *Z*-configuration at C-14/C-15. The coupling constants between H-6 and H-7 ( $J=15.4$  Hz), H-8 and H-9 ( $J=15.4$  Hz), and H-16 and H-17 ( $J=15.4$  Hz) indicated that the configurations at C-6/C-7, C-8/C-9, and C-16/C-17 are *trans*. HMBC correlations for H-20/C-4, H-21/C-10, and H-23/C-14 confirmed the methoxy substituents at these carbons. The existence of  $\alpha,\beta$ -unsaturated  $\delta$ -lactone ring was indicated by the chemical shift of H-5 ( $\delta$  4.98) implying the connectivity between O-5 and the carbonyl C-1 and the coupling constant for H-2/H-3 ( $J=10$  Hz)

**Fig. 2** Selected 2D NMR correlations and NOE observed for pterocidin (**1**).

that supports the *cis*-configuration for C-2/C-3. Thus, the structure of pterocidin was elucidated to be **1**. As for the stereochemistry, the relative configuration at C-4/C-5 was suggested to be *cis* by the strong NOESY correlation between H-4 and H-5. Absolute configuration of **1** is currently under investigation.

Pterocidin (**1**) is a new compound possessing an  $\alpha,\beta$ -unsaturated  $\gamma$ -oxygenated  $\delta$ -lactone at one end of the molecule with five chiral centers. Similar polyketide-derived  $\delta$ -lactones have been isolated from actinomycetes [8~12] and marine sponges [13, 14] and many of them show antitumor activity. The structure of **1** is closely related to sultricin [8], PD 113271 [9], and pironetin [10], antitumor compounds produced by *Streptomyces* species. Pterocidin (**1**) showed cytotoxicity against the cancer cell lines NCI-H522, OVCAR-3, SF539, and LOX-IMVI with  $\text{IC}_{50}$  values of 2.9, 3.9, 5.0, and 7.1  $\mu\text{M}$ , respectively, but no significant activity was exhibited against microorganisms.

## Experimental

### General Experimental Procedures

Optical rotation was recorded on a JASCO DIP-1030 polarimeter. The IR and UV spectra were taken on Shimadzu FT-IR300 and Hitachi U-3210 spectrophotometers, respectively. NMR experiments were performed on a JEOL JNM-LA400 NMR spectrometer with TMS as an internal standard. FAB mass spectra were obtained on a JEOL HX-110 spectrometer.

### Fermentation and Isolation

Isolation and fermentation of the producing strain *S. hygroscopicus* TP-A0451 was previously described [1, 15]. In brief, strain TP-A0451 was cultivated in a liquid medium, and the cultured whole broth (10 liters) was extracted with 1-butanol (10 liters). The organic layer was concentrated *in vacuo* to yield crude solid (33.7 g). The solid was suspended in  $\text{CH}_3\text{CN}$  and the soluble part was collected by filtration and evaporated to give dark brown oil

(6.6 g). This was chromatographed over silica gel column (CHCl<sub>3</sub>-MeOH=100:1~1:1) and the fraction eluted with CHCl<sub>3</sub>-MeOH=20:1 was collected and evaporated to give crude material (648 mg). It was further purified on C-18 column (200×40 mm, i.d., ODS-AM 120-S50, YMC Co., Ltd.) with a stepwise gradient of 20~80% CH<sub>3</sub>CN in 0.15% K<sub>2</sub>HPO<sub>4</sub> buffer (pH 3.5). Fractions containing pterocidin (eluted with 60% CH<sub>3</sub>CN in buffer) was evaporated *in vacuo* and the remaining aqueous solution was extracted with EtOAc. The organic layer was evaporated to dryness to yield pterocidin (11 mg).

### Cytotoxic Assay

IC<sub>50</sub> values of pterocidin against human cancer cell lines were determined in the same manner as described [16].

**Acknowledgments** We thank to Dr. T. Yamori at Cancer Chemotherapy Center, Japanese Foundation of Cancer Research for the evaluation of cytotoxicity of pterocidin.

## References

- Igarashi Y, Iida T, Yoshida R, Furumai T. Pteridic acids A and B, novel plant growth promoters with auxin-like activity from *Streptomyces hygrosopicus* TP-A0451. *J Antibiot* 55: 764–767 (2002)
- Sasaki T, Igarashi Y, Ogawa M, Furumai T. Identification of 6-prenylindole as an antifungal metabolite of *Streptomyces* sp. TP-A0595 and synthesis and bioactivity of 6-substituted indoles. *J Antibiot* 55: 1009–1012 (2002)
- Furumai T, Yamakawa T, Yoshida R, Igarashi Y. Clethramycin, a new inhibitor of pollen tube growth with antifungal activity from *Streptomyces hygrosopicus* TP-A0623. I. Screening, taxonomy, fermentation, isolation and biological properties. *J Antibiot* 56: 700–704 (2003)
- Achenbach H, Mühlenfeld A, Fauth U, Zähner H. Galbonolides A and B—two new non-glycosidic antifungal macrolides from *Streptomyces galbus*. *Tetrahedron Lett* 26: 6167–6170 (1985)
- Takatsu T, Nakayama H, Shimazu A, Furihata K, Ikeda K, Furihata K, Seto H, Otake N. Rustmicin, a new macrolide antibiotic active against wheat stem rust fungus. *J Antibiot* 38: 1806–1809 (1985)
- Ley SV, Neuhaus D, Williams DJ. A conformational study of elaiophyllin by X-ray crystallography and difference <sup>1</sup>H NMR method; observation of a selective sign reversal of the nuclear Overhauser effect. *Tetrahedron Lett* 23: 1207–1210 (1982)
- Shibata K, Satsumabayashi S, Nakagawa A, Ōmura S. The structure and cytotoxic activity of herbimycin C. *J Antibiot* 39: 1630–1633 (1986)
- Ohkuma H, Naruse N, Nishiyama Y, Tsuno T, Hoshino Y, Sawada Y, Konishi M, Oki T. Sultricin, a new antifungal and antitumor antibiotic from *Streptomyces roseiscleroticus*. Production, isolation, structure and biological activity. *J Antibiot* 45: 1239–1249 (1992)
- Hokanson GC, French JC. Novel antitumor agents CI-920, PD 113270, and PD 113271. 3. Structure determination. *J Org Chem* 50: 462–466 (1985)
- Kobayashi S, Tsuchiya K, Harada T, Nishide M, Kurokawa T, Nakagawa T, Shimada N, Kobayashi K. Pironetin, a novel plant growth regulator produced by *Streptomyces* sp. NK10958. *J Antibiot* 47: 697–702 (1994)
- Amemiya M, Someno T, Sawa R, Naganawa H, Ishizuka M, Takeuchi T. Cytostatin, a novel inhibitor of cell adhesion to components of extracellular matrix produced by *Streptomyces* sp. MJ654-NF4. II. Physico-chemical properties and structure determination. *J Antibiot* 47: 541–544 (1994)
- Hamamoto T, Seto H, Beppu T. Leptomycins A and B, new antifungal antibiotics. II. Structure elucidation. *J Antibiot* 36: 646–650 (1983)
- Kobayashi M, Higuchi K, Murakami N, Tajima H, Aoki S. Callystatin, a potent cytotoxic polyketide from the marine sponge, *Callyspongia truncata*. *Tetrahedron Lett* 38: 2859–2862 (1997)
- Sirirah S, Tanaka J, Ohtani I, Ichiba T, Rachmat R, Ueda K, Usui T, Osada H, Higa T. Bitungolides A~F, new polyketides from the Indonesian sponge *Theonella* cf. *swinhoei*. *J Nat Prod* 65: 1820–1823 (2002)
- Igarashi Y, Iida T, Sasaki T, Saito N, Yoshida R, Furumai T. Isolation of actinomycetes from live plants and evaluation of antiphytopathogenic activity of their metabolites. *Actinomycetol* 16: 9–13 (2002)
- Furumai T, Takagi K, Igarashi Y, Saito N, Oki T. Arisostatins A and B, new members of tetrocarcin class of antibiotics from *Micromonospora* sp. TP-A0316. I. Taxonomy, fermentation, isolation and biological properties. *J Antibiot* 53: 227–232 (2000)